

Karen Ferrante · Benjamin Winograd · Renzo Canetta

Promising new developments in cancer chemotherapy

Abstract The positive impact on survival of traditional chemotherapeutic agents has renewed interest in developing newer cytotoxic agents and orally active compounds with improved therapeutic indices. In addition, new insights into the pathways of human tumorigenesis have led to novel approaches aimed at specific mechanism-based targets. The taxane class, of which paclitaxel was the first member, has the unique ability to promote and stabilize microtubule function directly, thereby inhibiting mitotic progression and inducing apoptotic cell death. Paclitaxel provides treatment benefit in a broad range of solid tumors including breast, ovarian, and lung cancer. The success with paclitaxel stimulated interest in the microtubule as a new therapeutic target. Taxane analogues with improved preclinical efficacy have been identified and are entering clinical trials. The enthusiasm for oral anticancer agents and the therapeutic importance of platinum compounds has led to the development of JM216 (satraplatin), a novel platinum IV coordination complex with oral activity in cisplatin-resistant cell lines, which is now in phase III trials in prostate cancer. Another compound in late development is DPPE, a chemopotentiator that enhances the *in vivo* antitumor effects of cytotoxic agents such as doxorubicin, cyclophosphamide, and cisplatin. Agents that inhibit topoisomerase I and II have also been of interest. TAS-103 is a dual topoisomerase I and II inhibitor with pre-

clinical efficacy in a broad spectrum of tumors and in multidrug-resistant tumor cell lines. Vaccination strategies represent a rational therapeutic approach in the minimal residual disease or high-risk adjuvant therapy setting. The GMK and MGV vaccines utilizing ganglioside antigens overexpressed on human tumors such as melanoma and small cell lung cancer appear to induce antibody production reliably at tolerable doses and are under further clinical investigation. Inhibition of matrix metalloproteinases (MMPs) is another attractive target for intervention in several aspects of tumor progression. Local production of MMPs with subsequent degradation of the extracellular matrix is implicated in supporting tumor growth, invasion, and angiogenesis. The development of orally active, nontoxic MMP inhibitors is critical since these compounds will likely require chronic administration in conjunction with other therapies. Oncogenes and tumor suppressor genes are appealing targets for therapy since they are thought to be responsible for a significant number of cancers. Mutations in the Ras oncogene occur with great frequency in a number of human cancers including lung, pancreas, and colon cancer. Clinical development of potent and selective inhibitors of farnesyltransferase, the Ras-processing enzyme, is ongoing. These compounds uncouple Ras activity, affect tumor growth, and have demonstrated significant antitumor activity against experimental models of human cancer. The exciting compounds and novel therapeutic approaches currently under investigation by Bristol-Myers Squibb Pharmaceutical Research Institute offer great potential as effective cancer chemotherapy agents for the near future.

Work presented at the 14th Bristol-Myers Squibb Nagoya International Cancer Treatment Symposium, "Challenges in Cancer Metastasis," 11–12 September 1998, Nagoya, Japan

K. Ferrante · B. Winograd · R. Canetta
Bristol-Myers Squibb Pharmaceutical Research Institute,
Wallingford, CT, USA

K.J. Ferrante (✉)
Associate Director, Clinical Oncology,
Bristol-Myers Squibb Pharmaceutical Research Institute,
Richard L. Gelb Center for Pharmaceutical Research
and Development,
Wallingford, CT 06492-7660, USA
Tel.: +1 203 677 6384; Fax: +1 203 284 7690
e-mail: ferrantk@bms.com

Key words Cytotoxics · Taxanes · Vaccines · MMPI
Ras-FTI

Introduction

Traditional anticancer therapies have demonstrated the ability to prolong survival in the advanced disease set-

ting [35, 75, 78]. More importantly, they have influenced the primary therapy of cancer, where significant treatment benefits have been realized [5, 28, 62, 63, 66, 67]. The addition of chemotherapy to the treatment of patients with early-stage malignant disease improves the likelihood of long-term survival. Consequently, there has been a resurgence in interest in developing new cytotoxic agents, analogues, and orally active agents with altered or enhanced activity profiles. Additionally, recent research efforts have focused on novel mechanism-based approaches targeting pathways responsible for oncogenesis. This review discusses some of the innovative compounds under development by Bristol-Myers Squibb (BMS) Pharmaceutical Research Institute which offer promise for further substantial advancement in the successful treatment of human cancer.

Compounds in late development

Paclitaxel

The taxane class, of which paclitaxel was the first member, has the unique ability to promote and stabilize the assembly of tubulin into microtubules, the cellular structure that forms mitotic spindles and is required for chromosome segregation [20, 76]. In the presence of paclitaxel, cells accumulate in the mitotic phase of the cell cycle and undergo apoptosis as they attempt to divide. Paclitaxel is active and provides clinical benefit in a broad range of solid tumors including breast, ovarian, and lung cancer. Early clinical studies in advanced ovarian cancer demonstrated relevant antitumor activity and prolongation of time to progression in patients who had developed resistance to standard platinum-containing chemotherapy [24, 25, 49, 57, 84, 85]. Similarly, significant efficacy was observed with paclitaxel in breast cancer patients who had failed standard therapy, which included prior anthracyclines [1, 22, 65, 71]. The evidence for clinical non-cross-resistance between paclitaxel and the platinum or anthracyclines supported the evaluation of paclitaxel in combination with these important agents. Furthermore, it was logical to evaluate paclitaxel combination therapy in earlier stages of disease to maximize the potential beneficial impact of treatment.

In a randomized phase III trial conducted by the Gynecologic Oncology Group (GOG) in previously untreated patients with ovarian cancer, paclitaxel in combination with cisplatin resulted in significantly longer progression-free survival (median 18 vs 13 months, $P < 0.001$) and greater than one year longer median survival (median 38 vs 24 months, $P < 0.001$) compared with standard therapy with cyclophosphamide and cisplatin [58]. The results of the GOG-111 study have recently been confirmed by the European Organization for the Research and Treatment of Cancer (EORTC) Intergroup trial in advanced ovarian cancer: patients treated with the paclitaxel and cisplatin com-

bination had a significant advantage in terms of progression-free survival as well as overall survival compared to the cyclophosphamide and cisplatin treatment group [79]. Recently, Cancer and Leukemia Group B (CALGB) reported planned interim results from the phase III Intergroup study (CALGB 9344) that may represent the most significant advance in the adjuvant treatment of breast cancer in many years. This trial demonstrated that the sequential addition of paclitaxel to standard adjuvant therapy with doxorubicin and cyclophosphamide significantly prolonged the duration of disease-free and overall survival in women with node-positive breast cancer [36]. In terms of proportional benefit, paclitaxel reduced the recurrence rate by 22% and the death rate by 26%.

In advanced non-small cell lung cancer (NSCLC), the efficacy of paclitaxel in combination with cisplatin has been confirmed in three large phase III randomized controlled trials. Studies conducted by the Eastern Cooperative Oncology Group (ECOG), the EORTC, and BMS demonstrated superior response rates, increased time to progression, symptom alleviation, and comparable survival when paclitaxel in combination with cisplatin was compared to a cisplatin-containing regimen [6, 30, 31]. The activity of paclitaxel in advanced NSCLC was further confirmed in a recently completed phase III randomized trial in which single-agent paclitaxel significantly prolonged survival compared to best supportive care [83]. The full scope of potential therapeutic benefits from paclitaxel treatment, alone and in combination therapy, has yet to be realized and remains an active area of clinical research. In addition to studies in early-stage breast, ovarian, and lung cancer, there are ongoing trials in a wide range of tumor types including head and neck, bladder, small cell lung, endometrial, and gastric cancer.

JM-216 (satraplatin)

There is a renewed interest in oral chemotherapy drugs, given their ease of administration and potential for cost containment. JM-216 (satraplatin) (bis-acetato-ammine-dichloro cyclohexylamine platinum IV) is the first platinum analogue to be active by the oral route. In vitro cytotoxicity testing with JM-216 demonstrated comparable activity to cisplatin against several human and murine tumor cell lines, while cell lines resistant or innately insensitive to cisplatin did not show complete cross-resistance [41, 43, 69]. Generally, in vivo antitumor activity was comparable to that of parenterally administered cisplatin or carboplatin. JM-216 was ineffective in the few in vivo tumor models tested that were insensitive or resistant to cisplatin [41, 59, 73]. Combination studies in mice confirmed that JM-216 and oral etoposide were therapeutically synergistic in the intravenous P388 leukemia model [41].

Clinical studies with JM-216 were initiated in 1992 and to date approximately 400 patients have been

treated in phase I and II trials. Studies exploring different dosing schedules have demonstrated that the dose-limiting toxicities are myelosuppression (thrombocytopenia and neutropenia) and diarrhea [41]. Hematologic toxicity has been cumulative, requiring dose reduction and treatment delay, whereas nephrotoxicity and neurotoxicity have been infrequently reported. The recommended phase II dose is 100–120 mg/m² daily for 5 consecutive days every 3–4 weeks. In addition, a daily dose of 30–40 mg/m² for 14 consecutive days every 5 weeks is being further explored in phase II trials.

To date, activity with JM-216 has been demonstrated in phase II studies in small cell lung cancer, relapsed ovarian cancer, and hormone-refractory prostate cancer. Recently, a phase III trial has been initiated by the EORTC comparing JM-216 plus prednisone to prednisone alone in patients with hormone-refractory prostate cancer. The primary study endpoints are survival and time to pain progression. A second double-blind phase III trial in a similar population is planned in the USA and Australia. Phase I combination trials of JM-216 with several agents including oral etoposide, paclitaxel, uracil/ftorafur/tegafur, and leucovorin as well as with radiotherapy are ongoing.

DPPE

Compounds with the ability to improve the therapeutic index of currently available cytotoxic agents are under investigation. While DPPE was first synthesized >40 years ago as an antihistamine, it has recently been shown to act as a chemopotentiator enhancing the effects of doxorubicin, daunorubicin, carmustine, cyclophosphamide, and cisplatin while offering cytoprotection to the gut and normal bone marrow progenitor cells [11, 12, 23, 32, 50].

Although its mechanism of action has not been fully elucidated, DPPE is a potent antagonist for novel intracellular (microsomal and nuclear) histamine receptors (H_{1C}) which appear to be involved in the mediation of cellular proliferation [9]. Recent work suggests that DPPE may act as a reversal agent of the P-glycoprotein pump, as well as a mitochondrial poison [61].

An early phase I/II trial of DPPE in combination with various cytotoxic agents yielded evidence of activity in patients who had failed prior treatment with the cytotoxic agent alone [13]. More recently, impressive phase II results for DPPE in combination with doxorubicin have been reported by Brandes et al. in patients with metastatic breast cancer [10]. In 23 evaluable patients a response rate of 70% was observed (95% confidence interval [CI] 47–87%). The National Cancer Institute of Canada Clinical Trials Group (NCIC CTG) performed a confirmatory study [45]. In this trial, the overall response rate was 49% (95% CI 31–66%), which included 11% complete and 37% partial responses, which appears to be better than previously reported by the NCIC CTG for single-agent doxorubicin in a similar patient population

[68]. It is of note that in the NCIC CTG trial the observed toxicities were similar to those expected for single-agent doxorubicin with the exception of neurological effects attributable to DPPE. These effects included cerebellar signs and hallucinations that were managed with patient sedation during the DPPE infusion.

In view of these encouraging results, the NCIC CTG has initiated a phase III trial comparing the efficacy of doxorubicin plus DPPE to doxorubicin alone in metastatic breast cancer. In addition, phase II results for DPPE in combination with cyclophosphamide in hormone-refractory prostate cancer appear promising, with responses in measurable disease sites and prostate-specific antigen levels, as well as symptom palliation [23]. A randomized phase II trial of cyclophosphamide with and without DPPE in prostate cancer is underway.

Ganglioside vaccines

Progress in the identification of tumor-specific antigens and a better understanding of immune activation processes have led to new optimism for the development of successful cancer vaccines. Carbohydrate antigens abundantly expressed on the surface of most tumors serve as ideal targets for immune recognition and attack [27]. The GM2 and GD2 gangliosides are a class of carbohydrate antigens particularly well suited for cancer immunotherapy [8, 16, 18, 33, 64, 77, 90]. Ganglioside-based vaccines can effectively augment antibody responses against carbohydrate antigens and trigger the killing of antigen-positive tumor cells [53, 55], and there is evidence that vaccine-induced immune responses against a variety of tumor antigens are associated with a more favorable clinical outcome [42, 54]. In a double-blind trial, patients with stage III melanoma who were disease-free after surgery were randomized to receive treatment with the ganglioside GM2/bacille Calmette-Guérin (BCG) vaccine or BCG alone [54]. Vaccination with GM2/BCG was shown to induce primarily immunoglobulin M (IgM) antibodies in most patients. In addition, patients with preexisting or vaccine-induced GM2 antibodies had significantly longer disease-free and overall survival than those without an antibody response. Ganglioside-based vaccines are believed to be best suited for treatment in the adjuvant or minimal residual disease setting where the aim is to eradicate micrometastases and circulating tumor cells [53, 55].

The GM2-KLH/QS21 (GMK) and GM2-KLH/GD2-KLH/QS21 (MGV) vaccines are carbohydrate (ganglioside) vaccines conjugated to an immunogenic carrier protein, keyhole limpet hemocyanin, coadministered with the adjuvant QS-21. In a phase II trial, vaccination with GMK was well tolerated and appeared to induce anti-GM2 IgM antibodies consistently and at sustained levels [39]. The most common adverse events were grade I and II injection site reactions and mild transient fevers. The ECOG Intergroup is currently conducting a phase III trial comparing the relapse-free

and overall survival of patients with resected stage IIB and III melanoma randomized to treatment with either the GMK vaccine or interferon alfa-2b. The selection of the control arm was based on a previous ECOG study that demonstrated that interferon alfa-2b significantly increases relapse-free and overall survival in high-risk resected melanoma patients compared to observation [47]. The EORTC plans to perform a randomized trial of GMK versus observation in stage II melanoma patients.

Clinical studies with the MGv polyvalent ganglioside vaccine are also ongoing. In an initial phase I/II trial in melanoma patients conducted at the Memorial-Sloan Kettering Cancer Center, the MGv vaccine was shown to induce anti-GM2 and anti-GD2 antibody responses (both IgG and IgM) without significant vaccination-related toxicities [17]. Additional clinical trials will focus on tumors known to express both the GM2 and GD2 gangliosides such as small cell lung cancer; other tumor types are under study to determine their level and type of ganglioside expression. Large phase III randomized trials are required to establish the efficacy of the GMK and MGv vaccines because the vaccines are best suited for treatment in the minimal residual disease setting. Demonstration of prolongation of relapse-free and overall survival times are the study endpoints rather than eradication of well-established tumors.

Compounds in early development

Taxanes

The success of paclitaxel therapy in cancer treatment has stimulated interest in further improving its efficacy profile and in identifying new tubulin-active agents. Two taxane analogues (BMS-184476 and BMS-188797) demonstrate greater activity than paclitaxel or docetaxel in a number of rodent solid tumors as well as human xenografts, including one with multidrug resistance expression [81, 82]. In the distal site tumor models L2987 lung carcinoma and HCT/pk colon carcinoma both analogues were superior to paclitaxel by at least a 10-fold cell kill rate. In addition, BMS-184476 was superior to paclitaxel in the human xenograft A2780 ovarian carcinoma and showed modest improvements compared to docetaxel in the clinically derived HOC79 ovarian carcinoma cell line. BMS-188797 demonstrated superiority to paclitaxel in the M109 lung and HOC79 ovarian tumor models, and was moderately better in the A2780 ovarian distal tumor model. Both taxanes are presently in phase I trials evaluating different schedules of administration.

TAS-103

Topoisomerases (Topos) are nuclear enzymes with a critical role during the normal functioning of DNA synthesis and transcription. Currently available agents that target Topo I include topotecan and irinotecan.

Topo II inhibitors include etoposide, teniposide, amsacrine, and doxorubicin [29].

Recently, several compounds have been identified which inhibit both Topo I and Topo II [51, 70, 72]. TAS-103 is one such agent with a broad spectrum of antitumor activity and dual Topo inhibition in preclinical models [38, 80, 87]. In a standard assay, TAS-103 was equipotent to camptothecin and SN38 in inhibiting Topo I activity and 40-fold more potent than irinotecan [38, 80, 86]. In addition, in some model systems, TAS-103 was also more potent than etoposide in inhibiting Topo II activity. When compared to the dual Topo inhibitor intoplicine, TAS-103 was more potent in both Topo I and II assays [80].

Preclinical pharmacology studies have shown that TAS-103 has antitumor activity in a variety of animal and human tumor models and is non-cross-resistant with other Topo I or II inhibitors [2, 37, 80, 86, 87]. The *in vitro* cytotoxicity of TAS-103 was superior to that obtained with irinotecan, etoposide, and intoplicine in the P388 murine leukemia and human epidermoid carcinoma (KB) tumor cell lines. Interestingly, there was lack of cross-resistance *in vitro* with TAS-103 in cisplatin-, 5-fluorouracil-, and p-glycoprotein-resistant cells [2]. Additionally, there was partial and lack of cross-resistance in P388/CPT cells with decreased Topo I expression and KBNM4 cells with decreased Topo II expression, respectively. In a variety of *in vivo* studies, TAS-103 exhibited superior or comparable activity to concomitantly tested Topo I or Topo II inhibitors [2, 37, 80, 86, 87]. The models tested included the murine solid tumors B16-melanoma, colon 26, Lewis lung carcinoma, and Yoshida sarcoma model, as well as gastric, colon, breast, NSCLC, and pancreatic human tumor xenografts.

Two ongoing phase I trials with TAS-103 are evaluating different schedules of administration. The maximum tolerated dose of TAS-103 has yet to be defined in either trial and thus far the drug has been well tolerated.

Metalloproteinase inhibitors

Matrix metalloproteinases (MMPs) are a group of zinc-dependent proteolytic enzymes that are overexpressed in many pathological conditions including inflammatory disorders such as rheumatoid arthritis, osteoarthritis, multiple sclerosis, and cancer [4, 52]. In malignant disease, MMPs are capable of degrading the extracellular matrix, the principal barrier for tumor growth and metastasis [14, 88]. Thus MMP inhibitors (MMPIs) offer the potential to intervene in several aspects of tumor progression, including invasion, metastatic spread, and angiogenesis [14, 19, 88].

Malignant progression appears to be related to MMP activation and expression by tumors. Furthermore, the host plays an integral part in tumor growth and spread. There is evidence that host MMPs mediate the remodeling of extracellular matrix accompanying angiogenesis and that tumor cells can activate latent MMPs present in

host stromal tissue, contributing to metastasis [14, 19, 88]. Preclinical studies suggest that MMP-2 and MMP-9 are critical in the process of tumor cell invasion [88], and overexpression of these two MMPs has been shown to correlate with tumor aggressiveness and progression in retrospective studies. In tissue samples from NSCLC patients, a highly significant association was shown to exist between evidence of tumor spread and level of activated MMP-2 [15]. Additionally, elevated MMP-9 expression in surgical specimens from patients with colorectal cancer was linked with metastatic potential and was of prognostic significance for tumor recurrence and treatment outcome [89].

Tissue inhibitors of MMPs (TIMPs) are a class of specific natural protein inhibitors with the crucial role of keeping the damaging potential of the MMPs in check. An imbalance between the expression of activated enzyme and natural inhibitors involving greater MMP levels than TIMP levels has been shown to correlate with the degree of tissue destruction in malignant disease [14, 19, 88]. While the use of TIMPs as therapeutic agents would be impractical given their protein structure, the search for synthetic MMPi as a rational approach has been ongoing since the early 1980s [19].

BMS-275291 is an orally active MMPi in clinical development. *In vitro* studies have shown that BMS-275291 has potent activity against key MMPs including MMP-2 and MMP-9 [60]. Importantly, BMS-275291 does not inhibit the release of the tumor necrosis factor receptor nor does it cause joint toxicity in the marmoset monkey. These characteristics may have favorable implications in terms of the compound's expected clinical side effect profile. In particular, BMS-275291 has the potential to cause less joint toxicity than reported for other MMPi in development [21, 60, 88]. *In vivo*, oral BMS-275291 reduced the number and size of metastases of the rat HOSP-1 mammary tumor and was active against the murine B-16 melanoma tumor [60]. Furthermore, *in vitro* antiangiogenic assay results suggest that BMS-275291 may slow tumor growth by inhibiting angiogenesis. A phase I dose-escalation trial with BMS-275291 in healthy volunteers has been completed and studies in patients are planned.

Ras farnesyltransferase inhibitors

New understanding of two fundamental mechanisms of cancer, activation of protooncogenes and inactivation of tumor suppressor genes, has produced insight into the genetics of certain cancers. As a result, new biochemical targets have been identified for therapeutic use in anti-tumor therapy.

Among the oncogenes, activating mutations of the three functional Ras genes (H-Ras, K-Ras, and N-Ras) appear to occur at high frequency in a number of human cancers. The incidence of mutated Ras has been reported to varying degrees in different tumor types. For example, the incidence of mutated Ras in adenocarcinoma of the

pancreas, colon, and lung has been determined to be as high as 90%, 50%, and 30%, respectively [7]. The Ras protein plays a critical role in the transmission of growth signals from the external surface of a tumor cell to internal signaling pathways for regulation of cell proliferation and survival [3, 7, 56]. Mutated Ras proteins are locked in the persistently activated state, thereby contributing to the growth stimulation and malignant properties of cancer cells [3, 7, 44, 56]. The posttranslational modification of Ras, known as farnesylation, enables Ras proteins to anchor to the inner surface of the plasma membrane, which is critical for normal Ras function in signal transduction as well as for its oncogenic activities [44, 48]. Blocking farnesylation results in severe impairment in Ras function and uncoupling of the cell signal transduction cascade due to the inability of the nonfarnesylated protein to anchor to the cell membrane [26, 34, 40, 46, 74]. Thus Ras farnesyltransferase inhibitors (FTIs) should specifically target tumors dependent on oncogenic Ras.

BMS-214662 is a potent and selective FTI currently in clinical trials. When administered by the oral, intravenous, and intraperitoneal routes to tumor-bearing mice, BMS-214662 caused substantial regression of large established tumors [26]. *In vivo*, intravenous and oral BMS-214662 induced apoptosis in human tumor cells and was curative (cure rate > 50%) in a variety of human tumor xenograft models. Tumor regression of > 80% was observed in carcinogen-induced mouse tumors after treatment with BMS-214662. In addition, regression and cure were also seen in the highly resistant HCT-116/VM-26 human colon tumor variant, which is known to be paclitaxel insensitive. The preclinical efficacy data suggest that BMS-214662 has the potential to target human tumors with the H-Ras and K-Ras mutations as well as tumors without Ras mutations.

Conclusions

Intensive research has led to an explosion in the understanding of the molecular basis of carcinogenesis. Consequently, there is renewed interest in identifying new cytotoxic agents with improved or altered efficacy profiles, and there is greater focus on novel target-specific treatment approaches. Preclinical studies suggest that these new compounds have the ability to become effective anticancer therapies, and clinical trials are underway to evaluate the safety and efficacy of these agents. With such innovative and promising developments in cancer chemotherapy there is greater potential now than ever before to advance the treatment of malignant disease significantly.

References

1. Abrams JS, Vena DA, Baltz J, Adams J, Montello M, Christian M, Onetto N, Desmond-Hellmann S, Canetta R, Fried-

- man MA, Arbuick SG (1995) Paclitaxel activity in heavily pretreated breast cancer: a National Cancer Institute Treatment Referral Center trial. *J Clin Oncol* 13: 2056
2. Aoyagi Y, Utsugi I, Kobunai T, Yamada Y (1998) In vitro antitumor activity of a novel quinoline derivative, TAS-103, inhibiting topoisomerase I and II. *Proc Am Assoc Cancer Res* 39: A557
 3. Barbacid M (1987) Ras genes. *Ann Rev Biochem* 56: 779
 4. Baxter AD, Bird J, Bhogal R, Massil T, Minton KJ, Montana J, Owen DA (1997) A novel series of matrix metalloproteinase inhibitors for the treatment of inflammatory disorders. *Bioorg Med Chem Lett* 7: 897
 5. Bonadonna G, Brusamolino E, Valagussa P, Rossi A, Brugnatelli L, Brambilla C, De Lena M, Tancini G, Bajetta E, Musumeci R, Veronesi U (1976) Combination chemotherapy as an adjuvant treatment in operable breast cancer. *N Engl J Med* 294: 405
 6. Bonomi P, Kim K, Chang A, Johnson D (1996) Phase III trial comparing etoposide (E), cisplatin (C) versus taxol (T) with cisplatin G-CSF versus taxol-cisplatin in advanced nonsmall cell lung cancer. An Eastern Cooperative Oncology Group (ECOG) Trial. *Proc Am Soc Clin Oncol* 15: 382 (abstract)
 7. Bos JL (1989) Ras oncogenes in human cancer: a review. *Cancer Res* 49: 4682
 8. Bosslet K, Mennel HD, Rodden F, Bauer BL, Wagner F, Altmannsberger A, Sedlacek HH, Wiegandt H (1989) Monoclonal antibodies against epitopes on ganglioside GD2 and its lactones. *Cancer Immunol Immunother* 29: 171
 9. Brandes LJ, LaBelia FS (1993) Identification of intracellular histamine receptors (Hic) that regulate cell proliferation. *Adv Biosci* 89: 31
 10. Brandes LJ, Bracken SP (1998) The intracellular histamine antagonist, N,N-diethyl-2-[4(phenylmethyl)-phenoxy]ethanamine, HCl, may potentiate doxorubicin in the treatment of metastatic breast cancer: results of a pilot study. *Breast Cancer Res Treat* 49: 51
 11. Brandes LJ, LaBelia FS, Warrington RC (1991) Increased therapeutic index of antineoplastic drugs in combination with intracellular histamine antagonists. *J Natl Cancer Inst* 83: 1329
 12. Brandes LJ, McDonald KA, Bracken SP, Warrington RC (1993) Results of a human pilot study testing the hypothesis that the intracellular histamine antagonist DPPE increases the therapeutic index of doxorubicin. *Adv Biosci* 89: 375
 13. Brandes LJ, Simons KJ, Bracken SP, Warrington RC (1994) Results of a clinical trial in humans in refractory cancer of the intracellular histamine antagonist, N,N-diethyl-2-[4(phenylmethyl)phenoxy]ethanamine, HCl in combination with various single antineoplastic agents. *J Clin Oncol* 12: 1281
 14. Brown PD, Giavazzi R (1995) Matrix metalloproteinase inhibition: a review of antitumor activity. *Ann Oncol* 6: 967
 15. Brown PD, Bloxidge RE, Stuart NSA, Gatter KC, Carmichael J (1993) Association between expression of activated 72-kilodalton gelatinase and tumor spread in non-small cell lung carcinoma. *J Natl Cancer Inst* 85: 574
 16. Chang HR, Cordon-Cardo C, Houghton AN, Cheung NK, Brennan MF (1992) Expression of disialogangliosides GD2 and GD3 on human soft tissue sarcomas. *Cancer* 70: 633
 17. Chapman PB, Meyers ML, Williams L, Dantis L, Yao TJ, Israel R, Morrissey D, Hamilton WB, Houghton AN, Livingston PO (1998) Immunization of melanoma patients (pts) with a bivalent GM2/GD2 ganglioside conjugate vaccine. *Proc Am Assoc Cancer Res* 39: A2515 (abstract)
 18. Cheresch DA, Rosenberg J, Mujoo K, Hirschowitz L, Reisfeld RA (1986) Biosynthesis and expression of the disialoganglioside GD2, a relevant target antigen on small cell lung carcinoma for monoclonal antibody-mediated cytotoxicity. *Cancer Res* 46: 5112
 19. Davidson AH, Drummond AH, Galloway WA, Whittaker M (1997) Inhibition of matrix metalloproteinase enzymes. *Chem Industry* p. 258
 20. DeBrabander M, Gevens G, Nuydens R, Willebrords R, DeMey J (1981) Taxol induces the assembly of free microtubules in living cells and blocks the organizing capacity of the centrosomes and kinetochores. *Proc Natl Acad Sci USA* 78: 5608
 21. Denis LJ, Verweij J (1997) Matrix metalloproteinase inhibitors: present achievements and future prospects. *Invest New Drugs* 15: 175
 22. Dieras V, Marty M, Tubiana N, Corette L, Morvan F, Serin D, Mignot L, Chazard M, Garet F, Onetto N, Helmann S, Pouillart P (1995) Phase III randomized study of paclitaxel versus mitomycin in advanced breast cancer. *Semin Oncol* 22(suppl 8): 33
 23. DPPE Investigator Brochure (1997) Bristol-Myers Squibb Accession No. 910060959
 24. Einzig AI, Wlemik PH, Sasloff J, Runowicz DC, Goldberg GL (1992) Phase II study and long-term follow-up of patients treated with taxol for advanced ovarian adenocarcinoma. *J Clin Oncol* 10: 1748
 25. Eisenhauer EA, ten Bokkel Huinink WW, Swenerton KD, Gianni L, Myles J, van der Berg MEL, Kerr I, Vermorken JB, Buser K, Colombo N, Bacon M, Santabarbara P, Onetto N, Winograd B, Canetta R (1994) European-Canadian randomized trial of paclitaxel in relapsed ovarian cancer: high-dose versus low-dose and long versus short infusion. *J Clin Oncol* 12: 2654
 26. Farnesyltransferase Inhibitor (BMS-214662) Investigator Brochure (1998) Bristol-Myers Squibb Accession No. 91008582
 27. Feizi T (1985) Demonstration by monoclonal antibodies that carbohydrate structures of glycoproteins and glycolipids are onco-developmental antigens. *Nature* 314: 53
 28. Fisher B, Carbone P, Economou SG, Frelick R, Glass A, Lerner H, Redmond C, Zelen M, Band P, Katrych DL, Wolmark N, Fisher ER (1975) 1-Phenylalanine mustard (L-PAM) in the management of primary breast cancer: a report on early findings. *N Engl J Med* 292: 117
 29. Fisher DS, Knopf MT, Durivage HJ (eds) (1989). *The cancer chemotherapy handbook*, 4th edn. St. Louis: Mosby-Year Book, Inc.
 30. Gatzemeier U, von Pawel J, Gottfried M, ten Velde GPM, Mattson K, DeMarinis F, Harper P, Salvati F, Robinet G, Lucenti A, Bogaerts J, Winograd B, Gallant G (1998) Phase III comparative study of high-dose cisplatin (HD-cis) versus a combination of paclitaxel (TAX) and cisplatin (CIS) in patients with advanced non-small cell lung cancer (NSCLC). *Proc Am Soc Clin Oncol* 17: 1748A (abstract)
 31. Giaccone G, Splinter TAW, Debruyne C, Frelick R, Glass A, Lerner H, Redmond C, Zelen M, Band P, Katrych DL, Wolmark N, Fisher ER (1998) Randomized study of paclitaxel-cisplatin versus cisplatin-teniposide in patients with advanced non-small cell lung cancer. *J Clin Oncol* 16: 2133
 32. Glavin GB, Brandes LJ (1986) Antitumor and antisecretory effects of a novel diphenylmethane derivative and antiestrogen binding site ligand. *Can J Physiol Pharmacol* 66: 1139
 33. Hamilton WB, Helling F, Livingston PO (1993) Ganglioside expression on sarcoma and small-cell lung carcinoma compared to tumors of neuroectodermal origin. *Proc Am Assoc Cancer Res* 34: Abstract 2928
 34. Hancock JF, Magee AI, Childs JE, Marshall CJ (1989) All ras proteins are polyisoprenylated but only some are palmitoylated. *Cell* 57: 1167
 35. Henderson IC, Harris JR (1991) Principles in the management of metastatic disease. In: Harris JR, Hellman S, Henderson IC, Kinne DW (eds) *Breast diseases*, 2nd edn. Lippincott, Philadelphia, p 547
 36. Henderson IC, Berry D, Demetri G, Cirrincione L, Goldstein L, Martino S, Ingle JN, Cooper MR, Canellios G, Borden E, Fleming G, Holland JF, Graziano S, Carpenter J, Muss H, Norton L (1998) Improved disease-free (DFS) and overall survival (OS) from the addition of sequential paclitaxel (T) but not from the escalation of doxorubicin (A) dose level in the adjuvant chemotherapy of patients (PTS) with node-positive primary breast cancer (BC). *Proc Am Soc Clin Oncol* 17: 390A (abstract)

37. Hoshi A, Castaner J (1998) TAS-103, antineoplastic topoisomerase I and II inhibitor. *Drugs Future* 23: 513
38. Ishida T, Nishio K, Arioka H, Kurokawa H, Fukumoto H, Fukuoka K, Momoto T, Tomonari A, Yokote H, Iwamoto Y, Suzuki T, Usuda J, Saijo N (1997) Cytotoxic mechanisms of a novel DNA topoisomerase I and II dual inhibitor TAS-103. *Proc Am Assoc Cancer Res* 38: A137 (abstract)
39. Israel RJ, Chapman P, Myers M, Hamilton WB, Chang CH, Zhan C, Morrissey D, Livingston P (1998) Phase II clinical trial of GM2-KLH/QS-21 (GMK) vaccine in patients with malignant melanoma. 10th NCI-EORTC Symposium on New Drugs in Cancer Therapy; Abstract 322
40. Jackson JH, Cochrane CG, Boume JR, Solski PA, Buss JE, Der CJ (1990) Farnesol modification of Kirsten-ras exon 4B protein is essential for transformation. *Proc Natl Acad Sci USA* 87: 3042
41. JM216 Investigator Brochure (Version 2) (1998) Bristol-Myers Squibb Report Accession No.910068599
42. Jones PC, Sze LL, Llu PY, Morton DL, Irie RF (1981) Prolonged survival for melanoma patients with elevated IgM antibody to oncofetal antigen. *J Natl Cancer Inst* 66: 249
43. Kelland LR, Able G, McKeage MJ, Jones M, Goddard PM, Valenti M, Murrer BA, Harrap KR (1993) Preclinical antitumor evaluation of bis-acetato-ammine-dichloro-cyclohexylamine platinum (IV): an orally active platinum agent. *Cancer Res* 53: 2581
44. Kelloff GJ, Lubet RA, Fay JR, Steele VE, Boone CW, Crowell JA, Sigman CC (1997) Farnesyl protein transferase inhibitors as potential cancer chemopreventives. *Cancer Epidemiol Biomarkers Prev* 6: 267
45. Khoo K, Brandes L, Reyno L, Dent S, Vandenberg T, Lebowhl D, Fisher B, Eisenhauer E (1998) Phase II trial of DPPE and doxorubicin (DOX) chemotherapy in patients with metastatic breast cancer (BMC): A National Cancer Institute of Canada (NCIC) study. *Proc Am Soc Clin Oncol* 17: 583 (abstract)
46. Kim R, Rine J, Kim SH (1990) Prenylation of mammalian ras protein in xenopus oocytes. *Mol Cell Biol* 10: 5945
47. Kirkwood JM, Hunt Strawderman MH, Emstoff MS, Smith TJ, Borden EC, Blum RH (1996) Interferon alpha 2b adjuvant therapy of high-risk resected cutaneous melanoma: The Eastern Cooperative Oncology Group Trial EST 1684. *J Clin Oncol* 14: 7
48. Kohl NE, Wilson FR, Mosser SD, Giuliani E, DeSolms SJ, Conner MW, Anthony NJ, Holtz WJ, Gomez RP, Lee TJ, Smith RL, Graham SL, Hartman GD, Gibbs JB, Oliff A (1994) Protein farnesyltransferase inhibitors block the growth of ras-dependent tumors in nude mice. *Proc Natl Acad Sci USA* 91: 9141
49. Kohn EC, Sarosy G, Bicher A, Link C, Christian M, Steinberg SM, Rothenberg M, Adamo DO, Davis P, Ognibene FP, Cunnion RE, Reed E (1994) Dose-intense taxol: high response rate in patients with platinum-resistant recurrent ovarian cancer. *J Natl Cancer Inst* 86: 18
50. Kudoh K, Kikuchi Y, Hiramatsu H, Hirata J, Yamamoto K, Kita K, Nagata I (1997) Enhancement of antitumor activity of cisplatin by N,N-diethyl-2-[4-(phenylmethyl) phenoxy] ethanamine HCl in human ovarian cancer cells with intrinsic or acquired resistance to cisplatin. *Eur J Cancer* 33: 122
51. Larsen AK, Grondard L, Couprie J, Desioze B, Comoe L, Jardillier JC, Riou JF (1993) The antileukemic alkaloid fagarone is an inhibitor of DNA topoisomerases I and II. *Biochem Pharmacol* 46: 1403
52. Levy DE, Ezrin AM (1997) Matrix metalloproteinase inhibitor drugs. *Emerging Drugs* 2: 205
53. Livingston PO, Ragupathi G (1997) Carbohydrate vaccines that induce antibodies against cancer. 2. Previous experience and future plans. *Cancer Immunol Immunother* 45: 10
54. Livingston PO, Wong GYC, Adluri S, Tao Y, Padavan M, Parente R, Hanlon C, Jones Calves M, Helling F, Ritter G, Oettingen HF, Old LJ (1994) Improved survival in stage III melanoma patients with GM2 antibodies: a randomized trial of adjuvant vaccination with GM2 ganglioside. *J Clin Oncol* 12: 1036
55. Livingston PO, Zhang S, Lloyd KO (1997) Carbohydrate vaccines that induce antibodies against cancer. 1. Rationale. *Cancer Immunol Immunother* 45: 1
56. McCormick F (1989) Ras GTPase activating protein: signal transmitter and signal terminator. *Cell* 56: 5
57. McGuire WP, Rowinsky EK, Rosenshein NB (1989) Taxol: a unique antineoplastic agent with significant activity in advanced ovarian epithelial neoplasms. *Ann Intern Med* 111: 273
58. McGuire WP, Hoskins WJ, Brady MF, Kucera PR, Partridge EE, Look KY, Clarke-Pearson DL, Davidson M (1996) Cyclophosphamide and cisplatin compared with paclitaxel and cisplatin in patients with stage III and stage IV ovarian cancer. *N Engl J Med* 334: 1
59. McKeage MJ, Kelland LR, Boxall FE, Valenti MR, Jones M, Goddard PM, Gwynne J, Harrap KR (1994) Schedule dependency of orally administered bis-acetato-ammine-dichloro-cyclohexylamine-platinum (IV) (JM-216) in vivo. *Cancer Res* 54: 4118
60. Matrix Metalloproteinase Investigator's Brochure (Version 2) (1998) Bristol-Myers Squibb Accession No. 910068930
61. Menendez AT, Raventos-Suarez C, Fairchild C, Comell L, Lee F, Smykla R, Peterson R, Kramer R (1998) Mechanism of action of DPPE, a chemosensitizing agent. *Proc Am Assoc Cancer Res* 39: 3462 (abstract)
62. Moertel CG, Fleming TR, Macdonald JS, Haller DG, Laurie JA, Goodman PJ, Ungerleider JS, Emerson WA, Tormey DC, Glick JH, Veeder MH, Mailliard JA (1990) Lavarnisole and fluorouracil for adjuvant therapy of resected colon carcinoma. *N Engl J Med* 322: 352
63. Moertel CG, Fleming TR, Macdonald J, Haller D, Laurie JA, Tangen CM, Ungerleider JS, Emerson WA, Tormey DC, Glick JH, Veeder MH, Mailliard JA (1995) Fluorouracil plus levamisole as effective adjuvant therapy after resection of stage III colon carcinoma: a final report. *Ann Intern Med* 122: 321
64. Mujoo K, Cheresch DA, Yang HM, Reisfeld RA (1987) Disialoganglioside GD2 on human neuroblastoma cells: target antigen for monoclonal antibody-mediated cytotoxicity and suppression of tumor growth. *Cancer Res* 47: 1098
65. Nabholz JM, Gelmon K, Bontenbal M, Spielmann M, Cattel G, Conte P, Klaassen U, Namer M, Bonnetterre J, Fumoleau P, Winograd B (1996) Multicenter, randomized comparative study of two doses of paclitaxel in patients with metastatic breast cancer. *J Clin Oncol* 14: 1858
66. Nolvadex Adjuvant Trial Organisation (1985) Controlled trial of tamoxifen as a single adjuvant agent in the management of early breast cancer: analysis at six years by Nolvadex Adjuvant Trial Organisation. *Lancet* i: 836
67. Nolvadex Adjuvant Trial Organisation (1988) Controlled trial of tamoxifen as a single adjuvant agent in the management of early breast cancer: analysis at eight years by Nolvadex Adjuvant Trial Organisation. *Br J Cancer* 57: 608
68. Norris B, Pritchard K, James K, Myles J, Bennett K, Marlin S, Skillings J, Findlay B, Goss P, Latreille J, Lopez P, Osoba D, Rodgers A (1996) A phase III comparative study of vinorelbine (VNB) combined with doxorubicin (DOX) versus doxorubicin alone in metastatic/recurrent breast cancer (MBC): a National Cancer Institute of Canada (NCIC CTG) study. *Proc Am Soc Clin Oncol* 16: A59 (abstract)
69. Orr PM, O'Neill CF, Nicolson MC (1991) Platinum-resistant L1210 cell lines in new platinum drug development. In: SB Howell (ed), *Platinum and other metal coordination complexes in cancer chemotherapy*. New York, Plenum Publishing Co.; A69
70. Poddevin B, Riou J-F, Lavelle F, Pommier Y (1993) Dual topoisomerase I and II inhibition by intoplicine (RP-60475), a new antitumor agent in early clinical trials. *Mol Pharmacol* 44: 767
71. Reichman BS, Seidman AD, Crown JPA, Heelan R, Hakes TB, Lebowhl DE, Gilewski TA, Surbone A, Currie V, Hudis

- CA, Yao TJ, Klecker R, Jamis-Dow C, Collins J, Quinlivan S, Berkery R, Toomasi F, Canetta R, Fisherman J, Arbuck S, Norton L (1993) Paclitaxel and recombinant human granulocyte colony-stimulating factor as initial chemotherapy for metastatic breast cancer. *J Clin Oncol* 11: 1943
72. Riou J-F, Helissey P, Grondard L, Giorgi-Renault S (1991) Inhibition of eukaryotic DNA topoisomerase I and II activities by indoloquinolinedione derivatives. *Mol Pharmacol* 40: 699
 73. Rose WC, Crosswell AR, Schurig JE, Casazza AM (1993) Preclinical antitumor activity of orally administered platinum (IV) complexes. *Cancer Chemother Pharmacol* 32: 197
 74. Schafer WR, Kim R, Steme R, Thorner J, Kim SH, Rine J (1989) Generic and pharmacological suppression of oncogenic mutations in ras genes of yeast and humans. *Science* 245: 379
 75. Scheithauer W, Rosen H, Kornet GV, Sebesta C, Depisch D (1993) Randomised comparison of combination chemotherapy plus supportive care with supportive care alone in patients with metastatic colorectal cancer. *BMJ* 306: 752
 76. Schiff PB, Horwitz SB (1980) Taxol stabilizes microtubules in mouse fibroblast cells. *Proc Natl Acad Sci USA* 77: 1561
 77. Schulz G, Cheresch DA, Varki MN, Yu A, Staffileno LK, Reisfeld RA (1984) Detection of ganglioside GD2 in tumor tissues and sera of neuroblastoma patients. *Cancer Res* 44: 5914
 78. Stewart LA, Pignon JP (1995) Non-small Cell Lung Cancer Collaborative Group. Chemotherapy in non-small cell lung cancer: a meta-analysis using updated data on individual patients from 52 randomised clinical trials. *BMJ* 311: 899
 79. Stuart G, Bertelsen K, Mangioni C, Trope C, James K (1998) Updated analysis shows a slightly significantly improved overall survival (OS) for cisplatin-paclitaxel as first line treatment of advanced ovarian cancer: mature results of the EORTC-GCCG, NOCOVA, NCIC CTG and Scottish Inter-group trial. *Proc Am Soc Clin Oncol* 17: 1394 (abstract)
 80. TAS-103 Investigator's Brochure (1997) Taiho Pharmaceutical Co., Ltd.
 81. Taxane Analog (BMS-184476) Investigator Brochure (1997) Bristol-Myers Squibb Accession No. 910061274
 82. Taxane Analog (BMS-188797-01) Investigator Brochure (1998) Bristol-Myers Squibb Accession No. 910068570
 83. Thatcher N, Ranson M, Anderson H, Burt P, Davidson N, Nicolson M, Falk S, Carmichael J, Washington T, Jeynes A (1998) Phase III study of paclitaxel (TAXOL®) (T) versus best supportive care (BSC) in inoperable non-small cell lung cancer (NSCLC). ESMO 23rd Congress
 84. Thigpen JT, Blessing JA, Ball H, Hummel SJ, Barrett RJ (1994) Phase II trial of paclitaxel in patients with progressive ovarian carcinoma after platinum-based chemotherapy: A Gynecologic Oncology Group study. *J Clin Oncol* 12: 1748
 85. Trimble FL, Adams JD, Vena D, Hawkins MJ, Friedman MA, Fisherman JS, Christian MC, Canetta R, Onetto N, Hayn R, Arbuck S (1993) Paclitaxel for platinum-refractory ovarian cancer: results from the first 1,000 patients registered to National Cancer Institute Treatment Referral Center 9103. *J Clin Oncol* 11: 2405
 86. Utsugi T, Aoyagi K, Asao T, Okazaki S, Aoyagi Y, Sano M, Wierzba K, Yamada Y (1997) Antitumor activity of a novel quinoline derivative, TAS-103, with inhibitory effects on topoisomerases I and II. *Jpn J Cancer Res* 88: 992
 87. Utsugi T, Kobunai T, Aoyagi K, Aoyagi Y, Nakaoka M, Yamada Y (1997) Antitumor activity of TAS-103, a novel topoisomerase I and II inhibitor against orthotopically implanted human cancers. *Proc Am Assoc Cancer Res* 38: A2044 (abstract)
 88. Wojtowicz-Praga SM, Dickson RB, Hawkins MJ (1997) Matrix metalloproteinase inhibitors. *Invest New Drugs* 15: 61
 89. Zeng ZS, Huang Y, Cohen AM, Guillem JG (1996) Prediction of colorectal cancer relapse and survival via tissue RNA levels of matrix metalloproteinase-9. *J Clin Oncol* 14: 3133
 90. Zhang S, Cordon-Cardo C, Zhang HS, Reuter VE, Adluri S, Hamilton WB, Lloyd KO, Livingston PO (1997) Selection of tumor antigens as targets for immune attack using immunohistochemistry: 1. Focus on gangliosides. *Int J Cancer* 73: 42